

## Diterpenoid Alkaloids of *Delphinium buschianum* GROSSH.

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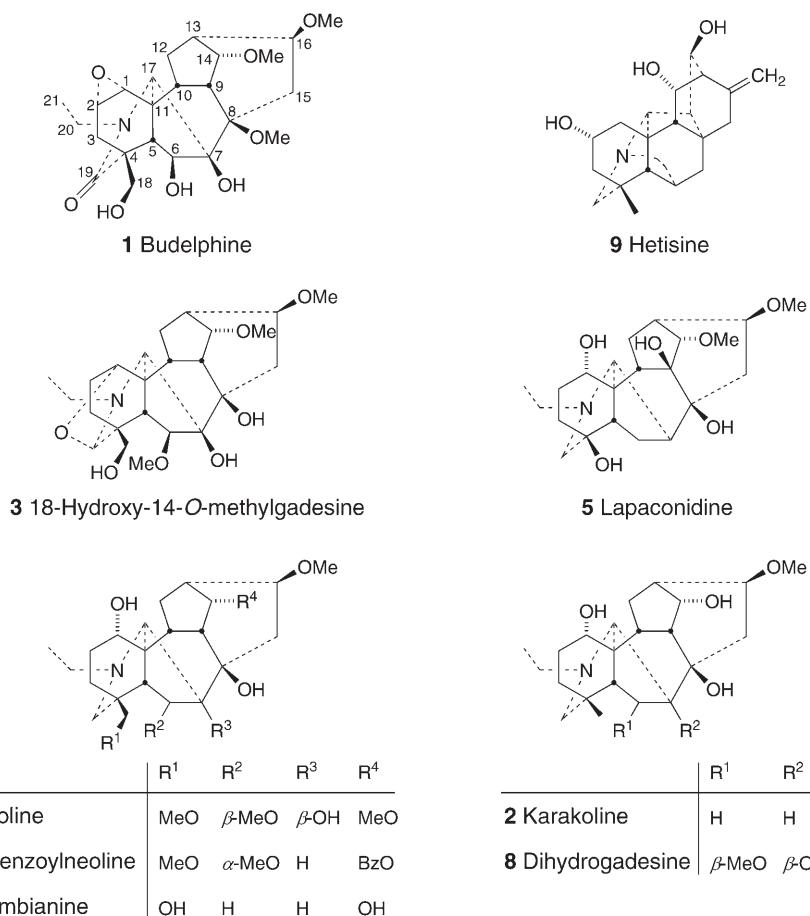
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From the aerial parts of *Delphinium buschianum* GROSSH., collected in Turkey, a new diterpenoid alkaloid **1**, named budelphine, was isolated along with the known diterpenoid alkaloids karakoline (**2**), 18-hydroxy-14-*O*-methylgadesine (**3**), delsoline (**4**), lapaconidine (**5**), columbianine (**6**), 14-benzoylneoline (**7**), and hetisine (**9**). The structure of **1** was established on the basis of <sup>1</sup>H-, <sup>13</sup>C-, DEPT, <sup>1</sup>H,<sup>1</sup>H-COSY, NOESY, HSQC, and HMBC NMR studies.

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**Introduction.** – *Delphinium* (larkspur) species are very toxic plants due to their diterpenoid alkaloid content. These alkaloids are neurotoxic agents, causing bradycardia, muscle-system spasms, hypotension, and death by arrest of respiration [1–3]. In continuation of our investigations on Turkish *Delphinium* species [4–8], we now report the alkaloid contents of *Delphinium buschianum* GROSSH. No previous work had been done on this species for its diterpenoid alkaloid constituents, except for the identification of methyllycaconitine obtained from this plant [9]. The chemical investigation of the aerial parts of *D. buschianum* led to the isolation of budelphine (**1**), a new diterpenoid alkaloid, together with karakoline (**2**), 18-hydroxy-14-*O*-methylgadesine (**3**), delsoline (**4**), lapaconidine (**5**), columbianine (**6**), 14-benzoylneoline (**7**), dihydrgadesine (**8**), and hetisine (**9**) [10–17].

**Results and Discussion.** – A novel diterpenoid alkaloid, designated as budelphine (**1**), was isolated from the aerial parts of *D. buschianum* collected at an altitude of 2100 m on Kızır Mountain, Kars-Arpaçay, Turkey. The molecular formula of **1**, C<sub>24</sub>H<sub>35</sub>NO<sub>8</sub> (EI-MS, M<sup>+</sup> at *m/z* 465), was derived by HR-MS (M<sup>+</sup> at *m/z* 465.54120; calc. 465.53950) and confirmed by the <sup>1</sup>H- and <sup>13</sup>C-NMR and DEPT data. The IR spectrum showed absorptions of OH groups and a lactam moiety at 3380 and 1656 cm<sup>−1</sup>, respectively, but no aromatic absorptions. The structure of **1** was established by its <sup>1</sup>H- and <sup>13</sup>C-NMR, DEPT, <sup>1</sup>H,<sup>1</sup>H-COSY, NOESY, HSQC, and HMBC data (*Tables 1* and *2*).



A broadband-decoupled  $^{13}\text{C}$ -NMR spectrum of **1** confirmed the presence of 24 C-atoms in the molecule. The DEPT spectra showed five quaternary C-atoms at  $\delta$  179.8, 86.8, 76.9, 48.3, and 37.9, ten signals for CH groups at  $\delta$  87.5, 83.7, 82.5, 80.4, 77.1, 67.0, 45.5, 44.6, 42.1, and 37.7, five signals for  $\text{CH}_2$  groups at  $\delta$  65.9, 50.0, 32.9, 29.6, and 24.6, and four signals for Me groups at  $\delta$  57.9, 57.6, 56.2, and 13.2. Diterpenoid alkaloids usually belong to either of two main groups, those with a  $\text{C}_{19}$  lycocitonine/aconitine-type skeleton with characteristic MeO groups and those derived from a  $\text{C}_{20}$  atisine-type one with an exocyclic methylene group [18]<sup>1</sup>). The  $^1\text{H}$ -NMR spectrum of budelphine (**1**) suggested the presence of three MeO groups; therefore, it should be a  $\text{C}_{19}$  norditerpenoid alkaloid, and according to the NMR signals there is an Et group attached to the N-atom ( $\delta(\text{C})$  13.2 (*q*) and 50.0 (*t*);  $\delta(\text{H})$  1.07 (*t*,  $J = 7$  Hz,  $\text{MeCH}_2\text{N}$ ) and 2.44–2.46 and 2.58–2.61 (*2m*,  $\text{MeCH}_2\text{N}$ )). The lactam =O group should be placed at C(19) with the signal at  $\delta(\text{C})$  179.8 (*s*) [19] and on the basis of the long-range  $^1\text{H}, ^{13}\text{C}$ -HMBC correlations

<sup>1</sup>) Lycocitonine = (1*a*,6*β*,14*a*,16*β*)-20-ethyl-4-(hydroxymethyl)-1,6,14,16-tetramethoxyaconitane-7,8-diol, aconitine = (1*a*,3*a*,6*a*,14*a*,15*a*,16*β*)-20-ethyl-1,6,16-trimethoxy-4-(methoxymethyl)aconitane-3,8,13,14,15-pentol 8-acetate 14-benzoate, and atisine = (6*R*,6*aR*,8*aS*,9*R*,11*S*,12*aR*,12*bS*,12*cS*)-decahydro-6-methyl-10-methylene-9*H*,12*cH*-8*a*,11-ethano-6,12*b*-propano-5*H*-benz[*h*]oxazolo[2,3-*a*]isoquinolin-9-ol.

Table 1.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data of Budelphine (**1**).  $\delta$  in ppm,  $J$  in Hz.

	$\delta$ (H)	$\delta$ (C)
$\text{H}_\beta\text{-C(1)}$	3.75 ( <i>d</i> , $J=9$ )	87.5 ( <i>d</i> )
$\text{H}_\beta\text{-C(2)}$	3.91–3.94 ( <i>m</i> )	82.5 ( <i>d</i> )
$\text{H}_a\text{-C(3)}$	1.73–1.77 ( <i>m</i> )	24.6 ( <i>t</i> )
$\text{H}_\beta\text{-C(3)}$	2.45 ( <i>dd</i> , $J=13, 5$ )	
C(4)	–	37.9 ( <i>s</i> )
H-C(5)	1.95–1.98 ( <i>m</i> )	37.7 ( <i>d</i> )
$\text{H}_a\text{-C(6)}$	4.40 (br. <i>s</i> )	77.1 ( <i>d</i> )
C(7)	–	86.8 ( <i>s</i> )
C(8)	–	76.9 ( <i>s</i> )
H-C(9)	1.79–1.81 ( <i>m</i> )	45.5 ( <i>d</i> )
H-C(10)	1.61–1.63 ( <i>m</i> )	42.1 ( <i>d</i> )
C(11)	–	48.3 ( <i>s</i> )
$\text{H}_a\text{-C(12)}$	2.29–2.31 ( <i>m</i> )	29.6 ( <i>t</i> )
$\text{H}_b\text{-C(12)}$	1.60–1.63 ( <i>m</i> )	
H-C(13)	2.39–2.41 ( <i>m</i> )	44.6 ( <i>d</i> )
H-C(14)	3.59 ( <i>t</i> , $J=5$ )	83.7 ( <i>d</i> )
$\text{H}_a\text{-C(15)}$	1.68–1.71 ( <i>m</i> )	32.9 ( <i>t</i> )
$\text{H}_b\text{-C(15)}$	2.54 ( <i>dd</i> , $J=12, 14$ )	
H-C(16)	3.75 ( <i>dd</i> , $J=7, 12$ )	80.4 ( <i>d</i> )
H-C(17)	2.88 ( <i>s</i> )	67.0 ( <i>d</i> )
$\text{H}_a\text{-C(18)}$	3.30 ( <i>d</i> , $J=10$ )	65.9 ( <i>t</i> )
$\text{H}_b\text{-C(18)}$	3.55 ( <i>d</i> , $J=10$ )	
C(19)	–	179.8 ( <i>s</i> )
$\text{H}_a\text{-C(20)}$	2.58–2.61 ( <i>m</i> )	50.0 ( <i>t</i> )
$\text{H}_b\text{-C(20)}$	2.44–2.46 ( <i>m</i> )	
Me(21)	1.07 ( <i>t</i> , $J=7$ )	13.2 ( <i>q</i> )
MeO-C(8)	3.42 ( <i>s</i> )	57.9 ( <i>q</i> )
MeO-C(14)	3.35 ( <i>s</i> )	57.6 ( <i>q</i> )
MeO-C(16)	3.32 ( <i>s</i> )	56.2 ( <i>q</i> )

of H-C(17) ( $\delta$ (H) 2.88 (*s*)) and  $\text{CH}_2$ (18) ( $\delta$ (H) 3.30 and 3.55 (each *d*,  $J=10$  Hz)). The three MeO groups ( $\delta$ (H) 3.32, 3.35, and 3.42 (*3s*);  $\delta$ (C) 56.2, 57.6, and 57.9 (*3q*)) could be positioned at CH(16) ( $\delta$ (H) 3.75 (*dd*,  $J=7, 12$  Hz);  $\delta$ (C) 80.4 (*d*)), CH(14) ( $\delta$ (H) 3.59 (*t*,  $J=5$  Hz);  $\delta$ (C) 83.7 (*d*)) and C(8) ( $\delta$ (C) 76.9 (*s*)), respectively [20]. According to the NMR and MS data, budelphine (**1**) should contain nine O-bearing C-atoms as shown by the signals at  $\delta$ (C) 179.8 (*s*), 87.5 (*d*), 86.8 (*s*), 83.7 (*d*), 82.5 (*d*), 80.4 (*d*), 77.1 (*d*), 76.9 (*s*), and 65.9 (*t*), but only eight O-atoms are present. Thus, besides the lactam group and three MeO groups, among the remaining five O-bearing C-atoms, three of them should carry OH groups, and an epoxy group should be placed between two C-atoms. The first two OH groups should be placed at C(7) ( $\delta$ (C) 86.8 (*s*)) and  $\text{CH}_2$ (18) ( $\delta$ (H) 3.30 and 3.58 (*2d*,  $J=10$  Hz);  $\delta$ (C) 65.9 (*t*)) [21], and the place of the third OH group should be at CH(6) in  $\beta$ -position ( $\delta$ (H) 4.40 (br. *s*);  $\delta$ (C) 77.1 (*d*)) [22]. Some norditerpenoid alkaloids contain an epoxy group between C(1) and C(19), but in **1**, C(19) is part of a lactam group. The epoxy group should be placed between CH(1) ( $\delta$ (H) 3.75 (*d*,  $J=9$  Hz);  $\delta$ (C) 87.5 (*d*)) and another C-atom; on the basis of the long range  $^1\text{H}$ ,  $^{13}\text{C}$ -HMBC correlations of H-C(5) ( $\delta$ (H) 1.95–1.98 (*m*)) and H-C(10) ( $\delta$ (H) 1.61–1.63 (*m*)) (Table 2) and NOESY correlations of H-C(1) ( $\delta$ (H) 3.20 (*d*,  $J=9$  Hz)) and H-C(5) ( $\delta$ (H) 1.95–1.98 (*m*)) (Table 2), the second C-atom should be CH(2) ( $\delta$ (H) 3.91–3.94 (*m*);  $\delta$ (C) 82.5 (*d*)).

Table 2. Summary of  $^1\text{H}$ ,  $^1\text{H-COSY}$ , NOESY, and HMBC Data of Budelphine (**1**)

	$^1\text{H}, ^1\text{H-COSY}$	NOESY	HMBC
$\text{H}_\beta-\text{C}(1)$	$\text{H}_\beta-\text{C}(2)$	$\text{H}_\beta-\text{C}(2), \text{H}-\text{C}(10), \text{H}_\text{b}-\text{C}(12)$	C(3), C(10)
$\text{H}_\beta-\text{C}(2)$	$\text{H}_\beta-\text{C}(1), \text{H}_\alpha-\text{C}(3)$	$\text{H}_\beta-\text{C}(1), \text{H}-\text{C}(5)$	C(5), C(10)
$\text{H}_\alpha-\text{C}(3)$	$\text{H}_\beta-\text{C}(2), \text{H}_\beta-\text{C}(3)$	–	–
$\text{H}_\beta-\text{C}(3)$	$\text{H}_\beta-\text{C}(2), \text{H}_\alpha-\text{C}(3)$	$\text{H}_\text{b}-\text{C}(18)$	C(1), C(2), C(19)
$\text{H}-\text{C}(5)$	$\text{H}-\text{C}(6), \text{H}-\text{C}(17)$	$\text{H}_\beta-\text{C}(2), \text{H}-\text{C}(6), \text{H}-\text{C}(9), \text{H}_\alpha-\text{C}(18)$	C(17), C(18), C(19)
$\text{H}_\alpha-\text{C}(6)$	$\text{H}-\text{C}(5)$	$\text{H}-\text{C}(5), \text{H}_\alpha-\text{C}(18), \text{H}_\text{b}-\text{C}(18)$	–
$\text{H}-\text{C}(9)$	$\text{H}-\text{C}(10), \text{H}-\text{C}(14)$	$\text{H}-\text{C}(5), \text{H}-\text{C}(10), \text{H}_\alpha-\text{C}(12), \text{H}-\text{C}(14)$	C(7), C(12), C(13), C(14), C(16)
$\text{H}-\text{C}(10)$	$\text{H}-\text{C}(9), \text{H}_\text{b}-\text{C}(12)$	$\text{H}_\beta-\text{C}(1), \text{H}-\text{C}(9), \text{H}_\alpha-\text{C}(12), \text{H}-\text{C}(14), \text{MeO}-\text{C}(16)$	–
$\text{H}_\alpha-\text{C}(12)$	$\text{H}_\text{b}-\text{C}(12), \text{H}-\text{C}(13)$	$\text{H}_\text{b}-\text{C}(12), \text{H}-\text{C}(13), \text{H}-\text{C}(14)$	–
$\text{H}_\text{b}-\text{C}(12)$	$\text{H}-\text{C}(10), \text{H}_\alpha-\text{C}(12)$	$\text{H}_\beta-\text{C}(1), \text{H}_\alpha-\text{C}(12), \text{H}-\text{C}(13), \text{H}-\text{C}(16), \text{H}-\text{C}(17)$	C(14), C(16)
$\text{H}-\text{C}(13)$	$\text{H}_\alpha-\text{C}(12), \text{H}-\text{C}(14)$	$\text{H}_\alpha-\text{C}(12), \text{H}_\text{b}-\text{C}(12), \text{H}-\text{C}(14), \text{H}-\text{C}(16), \text{MeO}-\text{C}(16)$	C(14), C(15), C(16)
$\text{H}-\text{C}(14)$	$\text{H}-\text{C}(9), \text{H}-\text{C}(13)$	$\text{H}-\text{C}(9), \text{H}-\text{C}(10), \text{H}_\alpha-\text{C}(12), \text{H}-\text{C}(13)$	C(16), MeO-C(14)
$\text{H}_\alpha-\text{C}(15)$	$\text{H}_\text{b}-\text{C}(15)$	$\text{H}-\text{C}(16), \text{MeO}-\text{C}(14)$	C(7), C(16)
$\text{H}_\text{b}-\text{C}(15)$	$\text{H}_\alpha-\text{C}(15), \text{H}-\text{C}(16)$	–	C(7), C(16)
$\text{H}-\text{C}(16)$	$\text{H}_\text{b}-\text{C}(15)$	$\text{H}_\text{b}-\text{C}(12), \text{H}-\text{C}(13), \text{H}_\alpha-\text{C}(15), \text{MeO}-\text{C}(16), \text{H}-\text{C}(17)$	C(12), C(14), MeO-C(16)
$\text{H}-\text{C}(17)$	$\text{H}-\text{C}(5)$	$\text{H}_\text{b}-\text{C}(12), \text{H}-\text{C}(16), \text{H}_\alpha-\text{C}(20), \text{H}_\text{b}-\text{C}(20), \text{Me}(21)$	C(5), C(7), C(10), C(19)
$\text{H}_\alpha-\text{C}(18)$	$\text{H}_\text{b}-\text{C}(18)$	$\text{H}-\text{C}(5), \text{H}-\text{C}(6), \text{H}_\text{b}-\text{C}(18)$	C(3), C(19)
$\text{H}_\text{b}-\text{C}(18)$	$\text{H}_\alpha-\text{C}(18)$	$\text{H}_\beta-\text{C}(3), \text{H}-\text{C}(6), \text{H}_\alpha-\text{C}(18)$	C(3), C(5), C(19)
$\text{H}_\alpha-\text{C}(20)$	$\text{H}_\text{b}-\text{C}(20), \text{Me}(21)$	$\text{H}-\text{C}(17), \text{H}_\text{b}-\text{C}(20), \text{Me}(21)$	C(17), C(19), C(21)
$\text{H}_\text{b}-\text{C}(20)$	$\text{H}_\alpha-\text{C}(20), \text{Me}(21)$	$\text{H}-\text{C}(17), \text{H}_\alpha-\text{C}(20), \text{Me}(21)$	C(17), C(19), C(21)
Me(21)	$\text{H}_\alpha-\text{C}(20), \text{H}_\text{b}-\text{C}(20)$	$\text{H}-\text{C}(17), \text{H}_\alpha-\text{C}(20), \text{H}_\text{b}-\text{C}(20)$	–
MeO-C(8)	–	–	C(8)
MeO-C(14)	–	$\text{H}_\alpha-\text{C}(15)$	C(14)
MeO-C(16)	–	$\text{H}-\text{C}(13), \text{H}-\text{C}(16)$	C(16)

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### Experimental Part

*General.* Vacuum liquid chromatography (VLC): *Merck Al<sub>2</sub>O<sub>3</sub>* (EM 1085) and SiO<sub>2</sub> 60 G (7731). Chromatographic separations: chromatotron with rotors coated with a 1 mm thick layer of *Merck Al<sub>2</sub>O<sub>3</sub>* 60 GF-254 (1092) or SiO<sub>2</sub> 60 PF-254 (7749). TLC: eluents toluene/AcOEt/Et<sub>3</sub>NH 7:4:1 or 7:4:2 and CHCl<sub>3</sub>/MeOH/NH<sub>4</sub>OH 5:3:1. Optical rotations: *Perkin-Elmer* 241 polarimeter. NMR Spectra: *Bruker* 500-MHz spectrometer. MS: *Finnigan MAT-90* spectrometer.

*Plant Material.* The aerial parts (500 g) of *Delphinium buschianum* GROSSH. were collected on Kizir Mountain Kars-Arpaçay, Turkey, at an elevation of 2100 m, in June 2002, and identified by one of us (H. Ö.). A voucher specimen was deposited in the Herbarium of the Faculty of Science and Literature, Süleyman Demirel University (No. Özçelik 9623), Isparta, Turkey.

*Extraction and Isolation.* The crude alkaloid extract (3.7 g) obtained from 1750 g of aerial parts was first separated by VLC (neutral Al<sub>2</sub>O<sub>3</sub>, petroleum ether/CHCl<sub>3</sub>/MeOH mixtures). The combined

Fractions 8 and 9 (with petroleum ether/CHCl<sub>3</sub> 60:40 → 55:45; 974 mg) were subjected to a SiO<sub>2</sub> rotor (petroleum ether/CHCl<sub>3</sub>/MeOH mixtures): delsoline (**4**; 17 mg), 18-hydroxy-14-O-methylgadesine (**3**; 22 mg), and lapaconidine (**5**, 14 mg). Fr. 10–13 (with petroleum ether/CHCl<sub>3</sub> 50:50 → 20:80; 437 mg) were subjected to a SiO<sub>2</sub> rotor (petroleum ether/CHCl<sub>3</sub>/MeOH mixtures): columbianine (**6**; 10 mg). Fr. 14 and 15 (with petroleum ether/CHCl<sub>3</sub> 10:90 → CHCl<sub>3</sub>; 375 mg) were subjected to a SiO<sub>2</sub> rotor (petroleum ether/CHCl<sub>3</sub>/MeOH mixtures): karakoline (**2**; 50 mg) and budelphine (**1**; 18 mg;  $[\alpha]_D^{20} = +6.95$  ( $c = 0.18$ , CHCl<sub>3</sub>)). Fr. 18–20 (with CHCl<sub>3</sub>/MeOH 96:4 → 92:8; 736 mg) were subjected to a SiO<sub>2</sub> rotor (petroleum ether/CHCl<sub>3</sub>/MeOH mixtures): 14-benzoylneoline (**7**; 11 mg) and dihydrogadesine (**8**; 48 mg). Fr. 22–28 (with CHCl<sub>3</sub>/MeOH 80:20 → MeOH; 303 mg) were subjected to a Al<sub>2</sub>O<sub>3</sub> rotor (petroleum ether/CHCl<sub>3</sub>/MeOH mixtures): hetisine (**9**; 25 mg). Delsoline (**4**), lapaconidine (**5**), karakoline (**2**), and hetisine (**9**) were obtained in pure state directly from chromatotron fractions.

Budelphine (=*(1α,2α,6β,14α,16β)-1,2-Epoxy-20-ethyl-6,7-dihydroxy-4-(hydroxymethyl)-8,14,16-trimethoxyaconitan-19-one*; **1**), **7**, **8**, and **3** were purified by prep. TLC (SiO<sub>2</sub>, toluene/AcOEt/Et<sub>2</sub>NH 7:4:1) and **6** by prep. TLC (SiO<sub>2</sub>, toluene/AcOEt/Et<sub>2</sub>NH 7:4:2). All known compounds were identified by comparison of their <sup>1</sup>H- and <sup>13</sup>C-NMR data and co-TLC behavior with those of authentic samples.

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